



E3scan™ Ligand Binding Assay Platform and SPRINTer™ Cell Lines for Targeted Protein Degradation and PROTAC® Discovery

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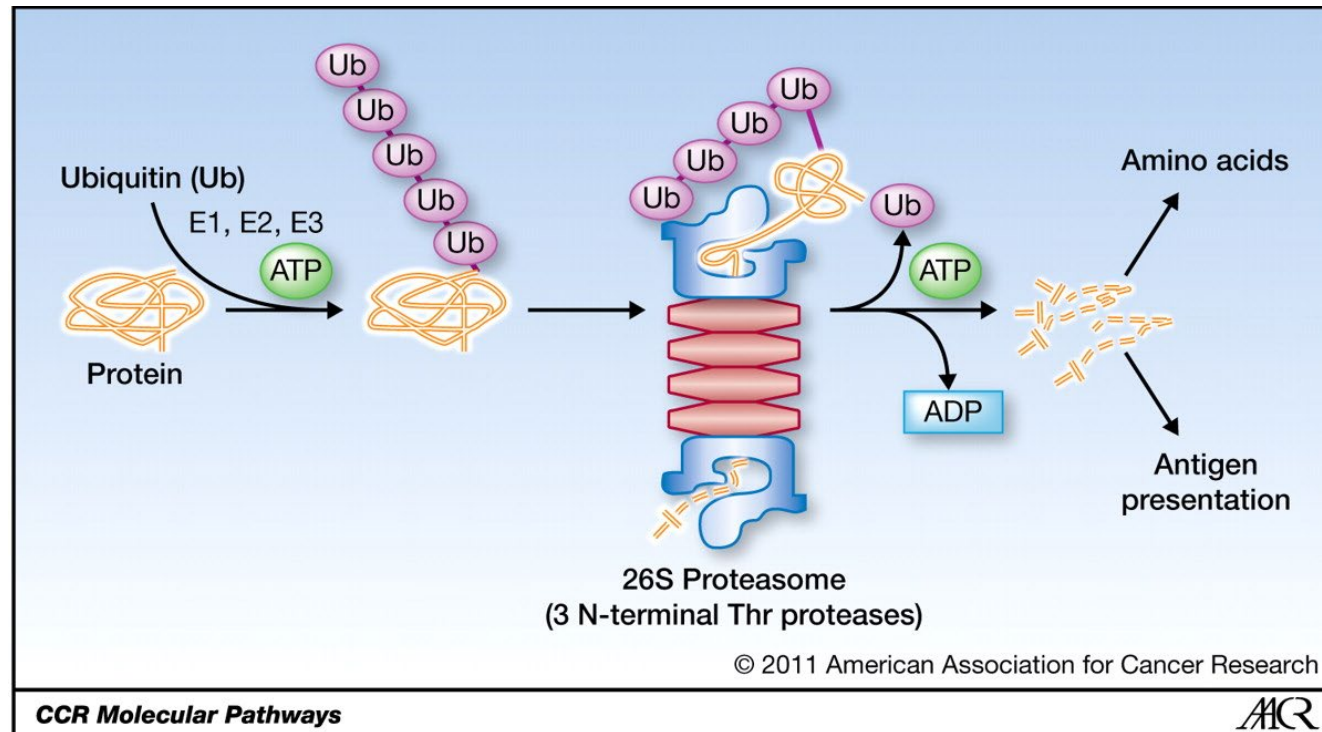
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**A STRONG
FOUNDATION
FOR SUCCESSFUL
DRUG DISCOVERY**

- Eurofins Discovery Targeted Protein Degradation Solutions Overview
- E3scan™ Technology and Assay Validation
 - CRBN with DDB1, VHL with Elongin B/C and other examples
- SPRINTer™ Biosensor Cell Lines For Screening Applications
- SPRINTer Assay Characterization
- Summary

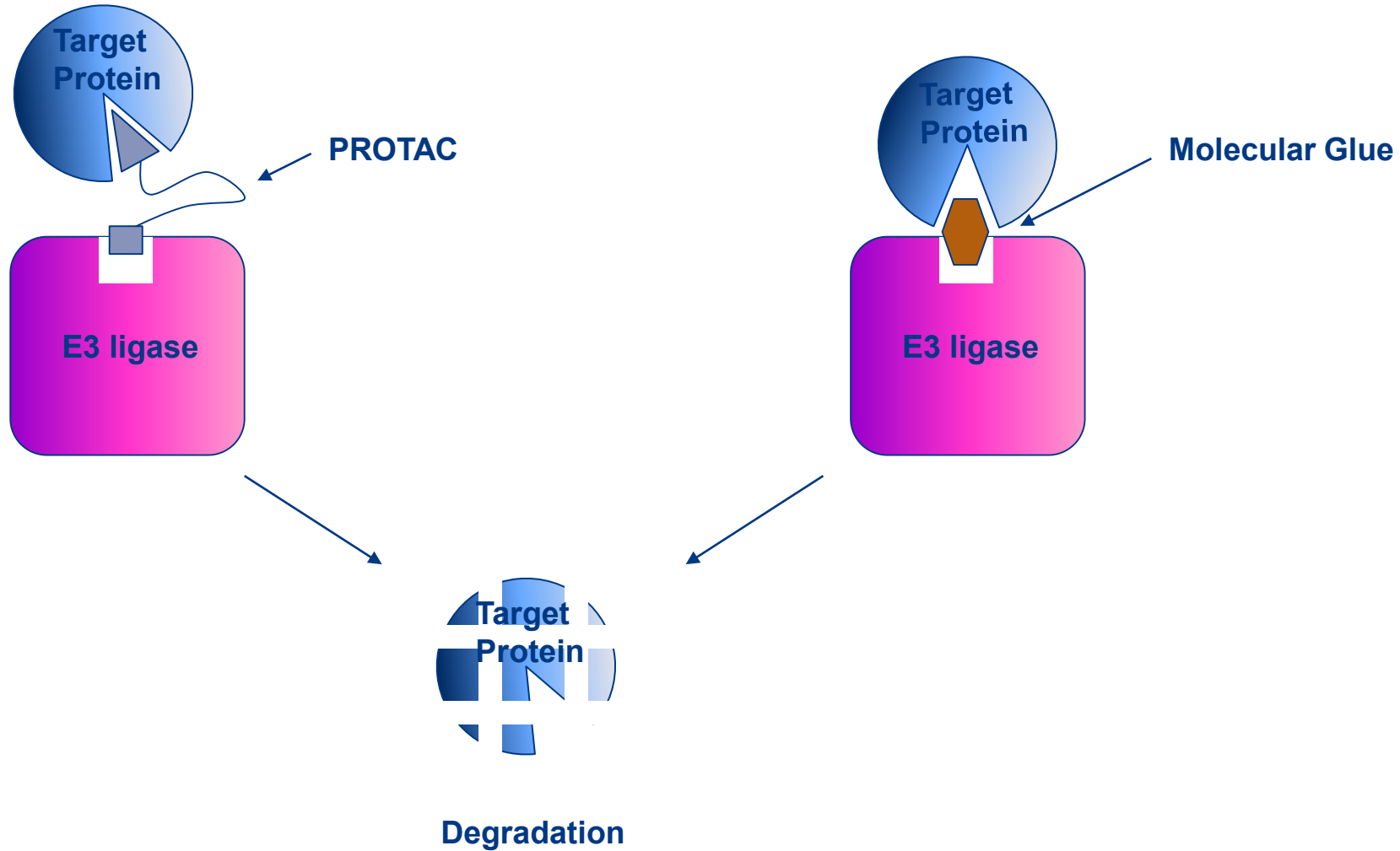
Targeted Protein Degradation

The ubiquitin-proteasome system controls protein levels by tagging proteins with poly-ubiquitin (on specific lysine residues), which results in their degradation by the proteasome



Goal: To manipulate the endogenous ubiquitin–proteasome system to achieve targeted degradation of specific proteins within cells

Approaches in Targeted Protein Degradation: PROTAC[®] - Proteolysis-Targeting Chimera, and Molecular Glue



PROTAC is a trademark of Arvinas

Chemistry
(Beyond Rule of 5)

Cell-Based Assays and
Phenotypic Platforms

Biochemical Platforms

Biochemical Assays

E3scan™ ligand binding
assay

Binding to E3 Ligase

KINOMEscan®,
Bromoscan®, BCL2scan™

Binding to Target
Protein or Disease
Protein

SPRINTer™ cell lines
(to assess target
degradation)

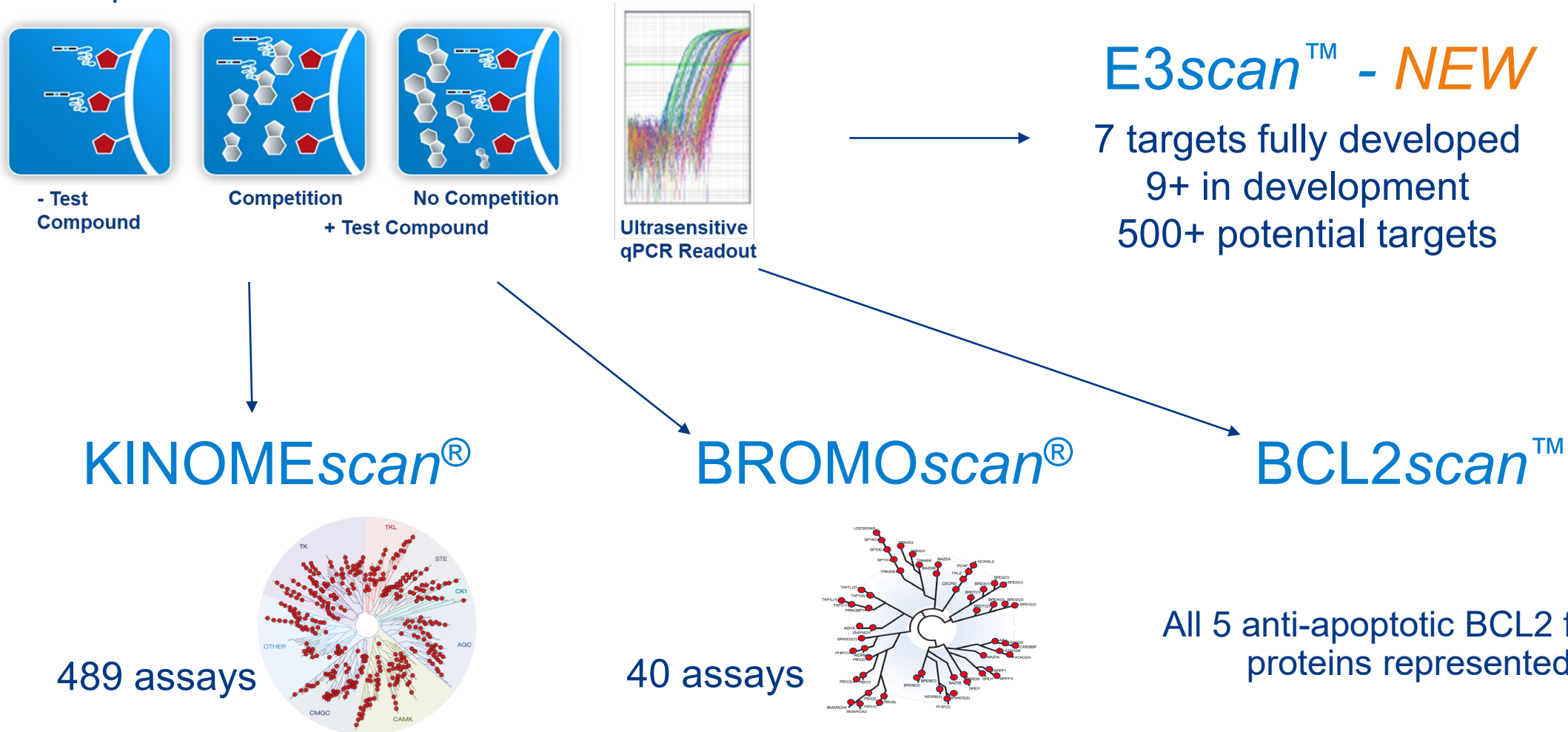
InCELL Target
Engagement Assays (for
E3 ligase or Target)

Phenotypic approaches
using the BioMAP®
Platform (for profiling
and screening)

Synthetic Chemistry
Medicinal Chemistry

Utilizing Novel Family-Wide Screening and Profiling Technology

We can utilize our unique competition binding assays using DNA-tagged protein targets: qPCR readout. Protein purification not required



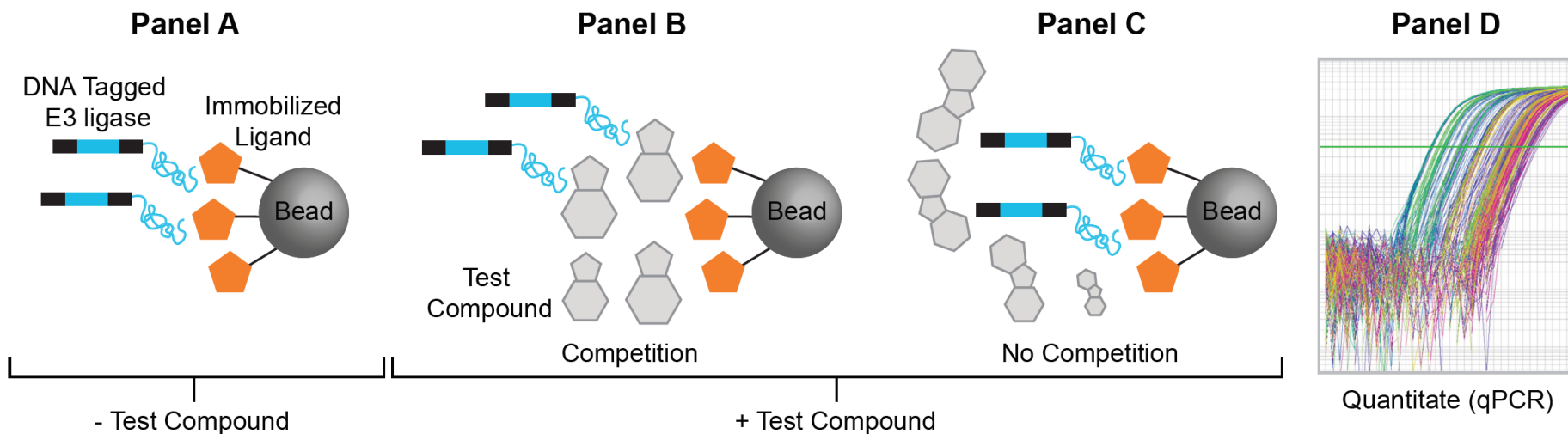
E3scan™ Assay Principle Using KINOMEscan® Platform

Ligand binding site-directed competition binding assays

Three key components

- E3 ligase tagged with DNA (low pM E3 ligase concentration in assay)
 - Expression in mammalian cells or by using proprietary T7 phage display system
- Known E3 ligase ligand (small molecule or peptide) immobilized on solid support
- Test compound or solvent control

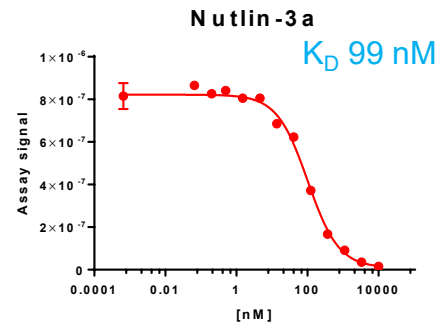
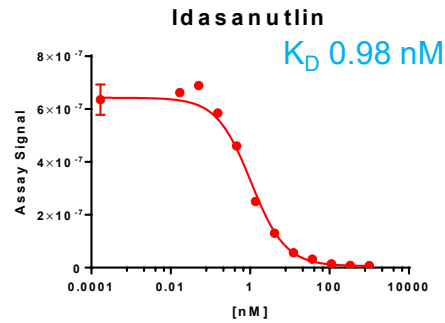
Measure amount of E3 ligase captured by solid support in the presence or absence of a test compound (ultrasensitive qPCR readout)



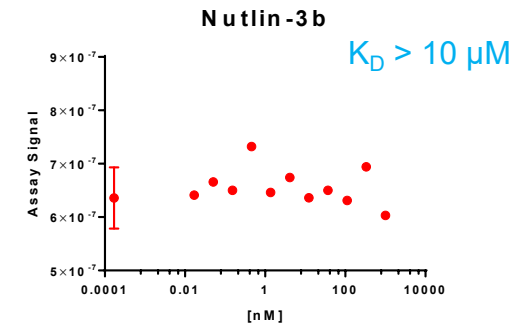
Full Length MDM2 and MDMX E3scan™ Assay Validation Data

MDM2

MDM2 Positive controls:

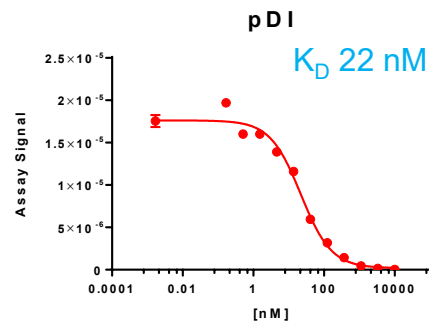


MDM2 Negative control:

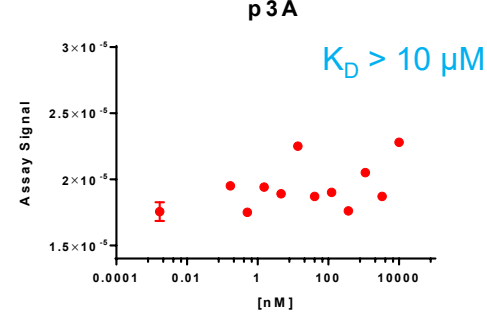


MDMX

MDMX Positive controls



MDMX Negative control:



MDM2 and MDMX assay validation

- Robust assay (assay window >50)
- Accurate data for control compound

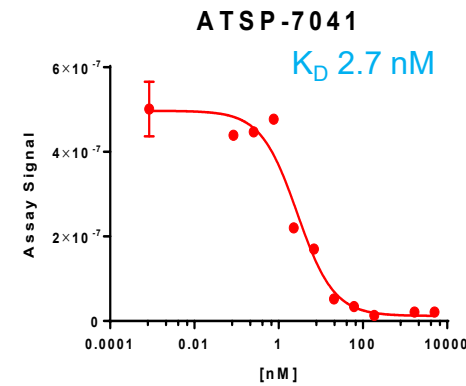
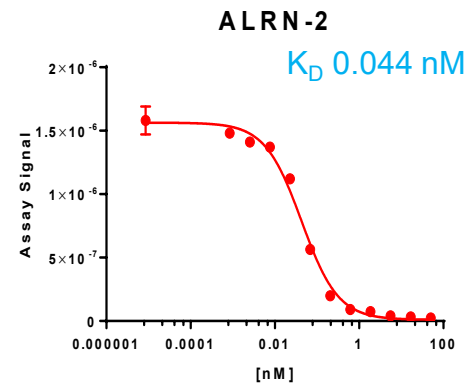
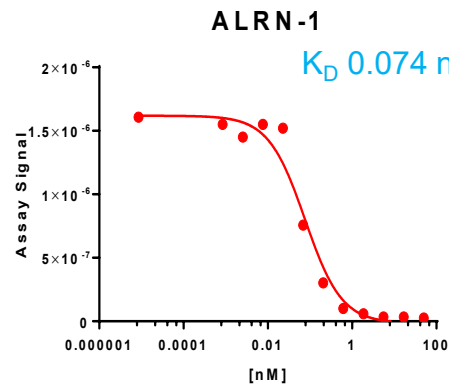
Compound Name	K_D (nM)	Literature value
Idasanutlin	0.98	6 nM by HTRF ¹
Nutlin-3a	99	90 nM by SPR ²
Nutlin-3b	>10,000	13.6 μM by SPR ²
pDI	17	100 nM by ELISA ³
p3A	>10,000	Non detected by ELISA ³

1. Ding et al, 2013
2. Vassilev et al, 2004
3. Hu et al, 2007

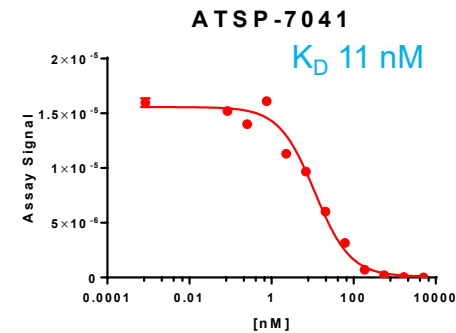
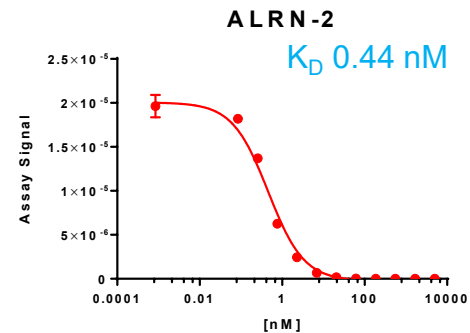
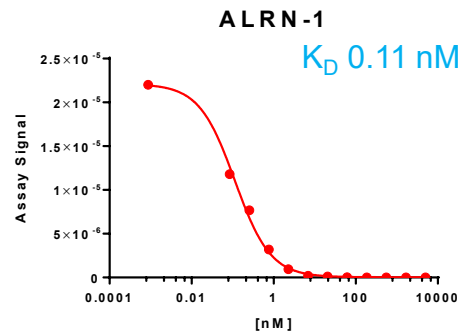
MDM2 vs. MDMX E3scan™ Assay Validation Data - Aileron High-Affinity Stapled Peptides



MDM2:



MDMX:

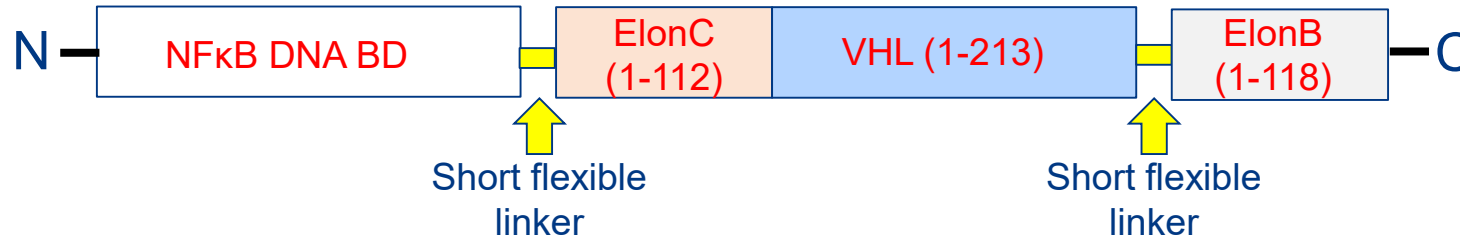


K_D measurements for the interactions of Aileron's stapled peptides with MDM2 and MDMX.

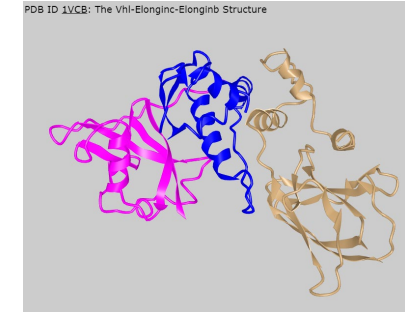
- The measured K_D s for ATSP-7041 stapled peptide are comparable to literature values measured using Biacore, 0.91 and 2.31 nM, respectively (Chang YS, et al. *PNAS*, 2013).

VHL (elongin BC) E3scan™ Assay Validation Data

VHL fused to ElonC (NP_005639.1) and ElonB (NP_009039.1) – all full-length

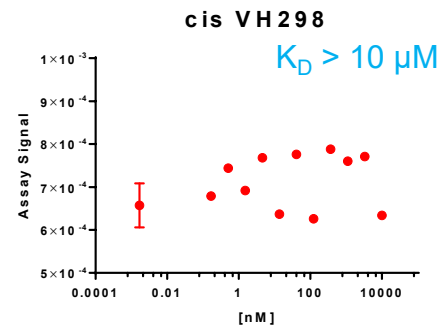
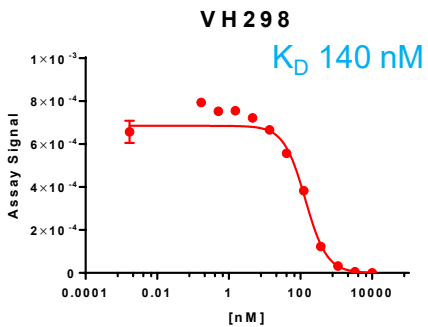


Inspired by
PDB 1VCB



Positive control:

Negative control:

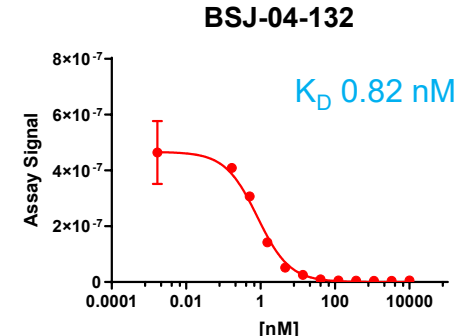
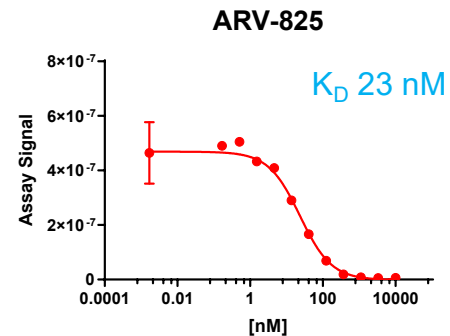
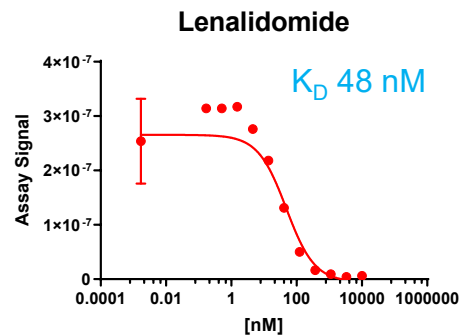


E3 ligase	Compound	AVG K_D (nM)	AVG Assay Window
VHL (elongin BC)	VH298	130	840
VHL (elongin BC)	Cis VH298	>10,000	0.9

VHL (elongin BC) assay validation

- Robust assay (assay window >800)
- Accurate data for VH298, compare to 80-90 nM by FP and ITC (Frost J, et al. *Nat Commun*, 2016) and negative control Cis VH298 (no binding)
- Additional validation from compounds that cannot be disclosed

Full Length CRBN E3scan™ Assay Validation Data



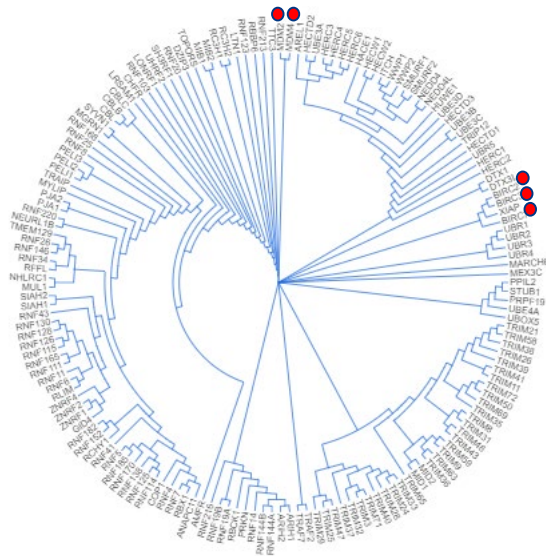
CRBN assay validation

- Robust assay (assay windows >30)
- Accurate rank order K_D s for three control inhibitor compounds: Pomalidomide, Lenalidomide, Thalidomide (literature values obtained by FP are 157, 178, 250 nM respectively, Fischer ES, et al. Nature, 2014)
- Co-expression with DDB1 does not impact inhibitors K_D s and assay signals
- No binding detected for matching negative control PROTACs

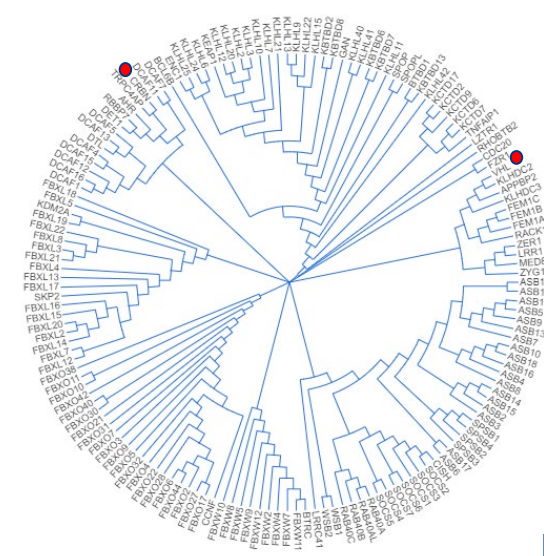
Compound Name	CRBN AVG K_D (nM)	CRBN + DDB1 AVG K_D (nM)	Compound Name	CRBN AVG K_D (nM)	CRBN + DDB1 AVG K_D (nM)
ARV-825	37	22	Lenalidomide	24	31
BSJ-03-123	4.1	4.0	Pomalidomide	20	22
BSJ-03-204	1.4	1.4	Thalidomide	48	44
BSJ-04-132	1.3	0.77	THAL SNS 032	18	13
dBET1	3.8	4.8	TL 12-186	10	5.7
dBRD9	3.6	4.0	TL 13-112	18	11
dTAG-13	113	93	ZXH 3-26	2.9	1.6

- We have developed and validated E3scan assays against MDM2, MDMX, VHL, CRBN, cIAP1, cIAP2, and XIAP
- All assays are robust and high throughput and give high quality K_D curves
- Correct potency and rank order for the control inhibitors tested
- Assays do not approach the tight binding limit – even for pM compounds and stapled peptides
- More E3 targets in development, not limited by available small molecule ligand

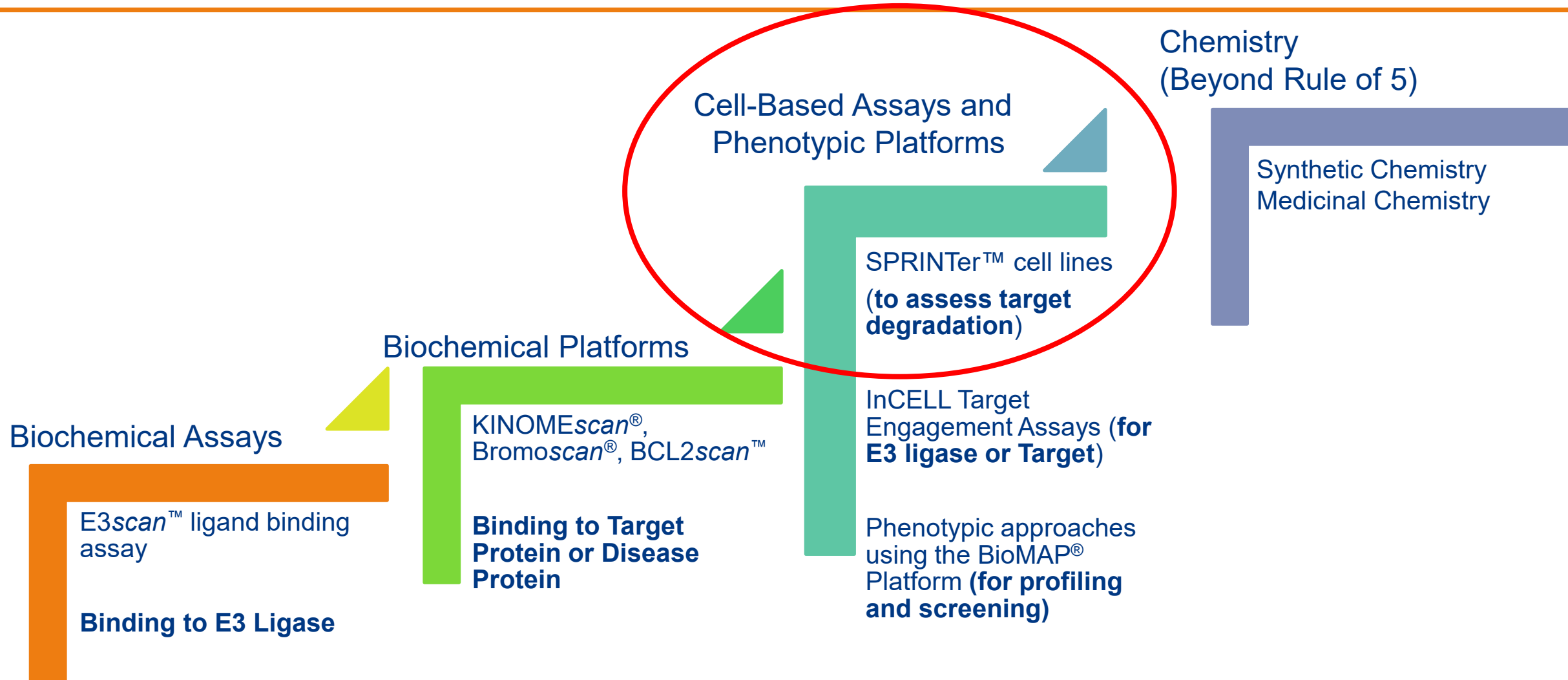
Simple E3 ligases tree



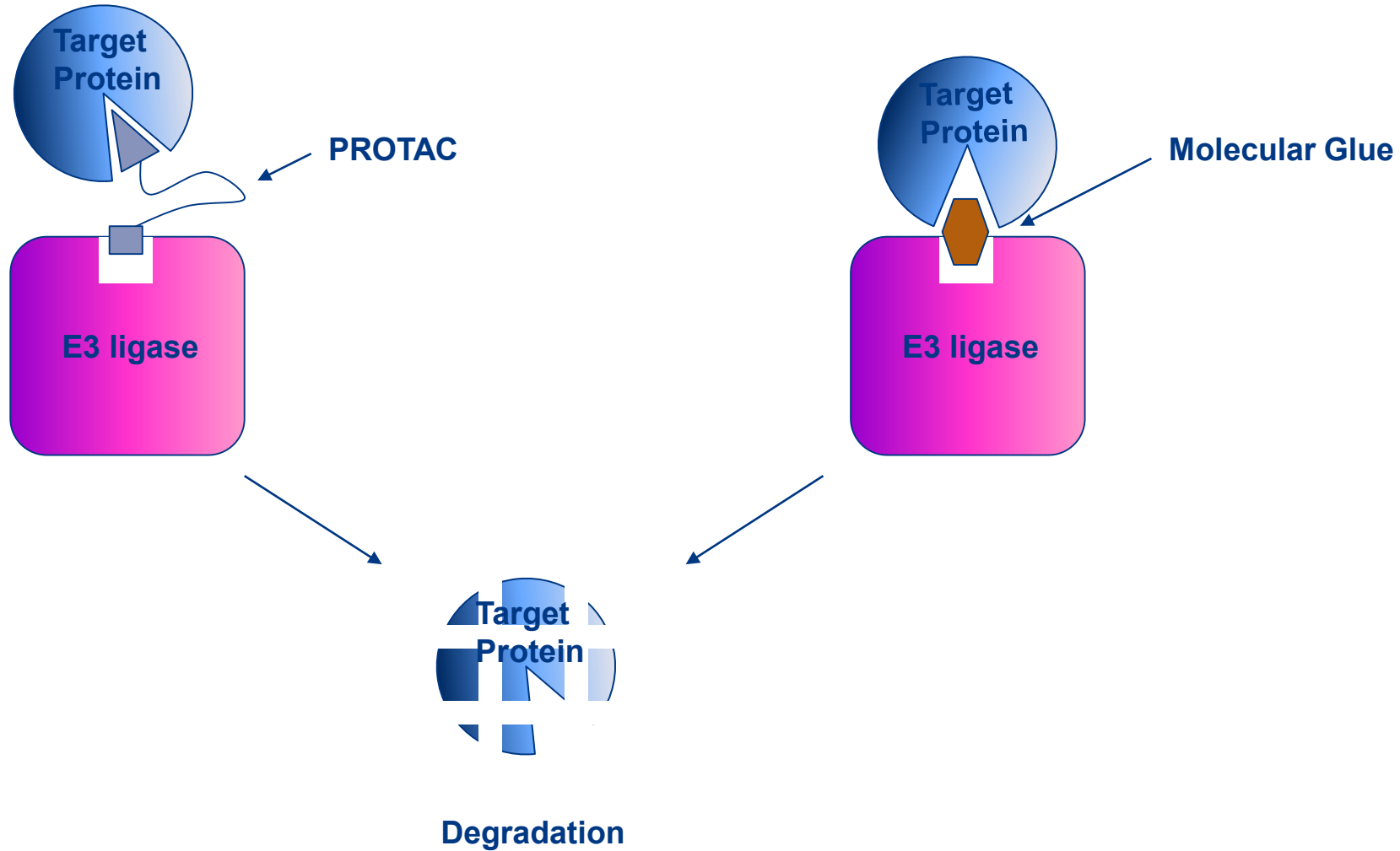
Complex E3 ligases tree



<https://ubihub.thesgc.org/static/UbiHub.html>



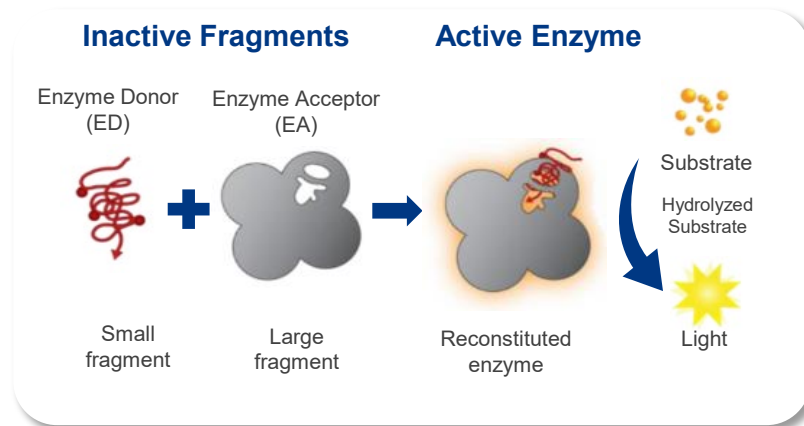
Approaches in Targeted Protein Degradation: PROTAC[®] - Proteolysis-Targeting Chimera, and Molecular Glue



PROTAC is a trademark of Arvinas

Evaluating On-Target Activity of Targeted Degradator Molecules

- Compatible with disease cell models expressing physiologically relevant levels of the target protein and relevant E3 ligase(s)
 - Over-expression systems incompatible with certain degrader molecules, such as PROTACs
- A sensitive and scalable method for detection of protein turnover
 - Commonly utilized techniques include Westerns (low-throughput), ELISAs, or proliferation assays
- Robust, reproducible, and easy to implement

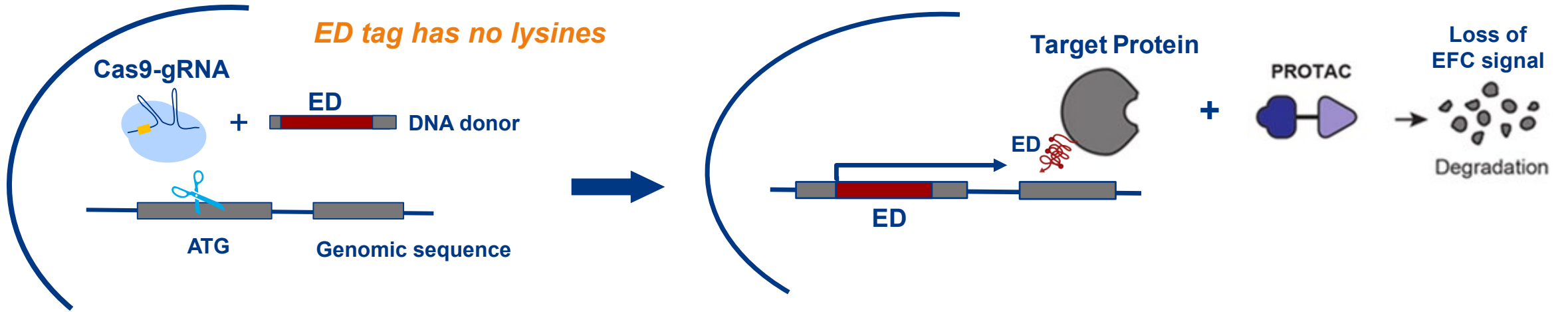


Solution:

Enzyme Fragment Complementation (EFC)

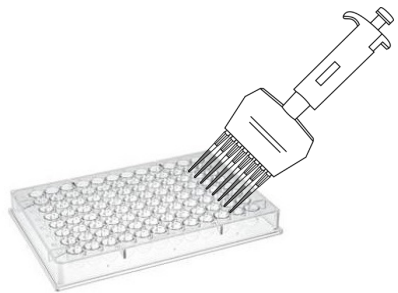
Combines FM reporter sensitivity with ease of implementation (simple, homogeneous assay format)

Assay Concept: Employ CRISPR / Cas9 to Introduce ED Tag into Endogenous Target

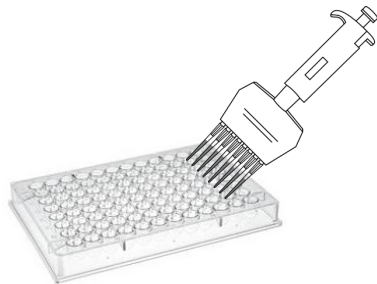


SPRINTer™ Assay Design: A Simple, Rapid and Homogeneous Assay Workflow

Seed stable biosensor cells in 96- or 384-well plate

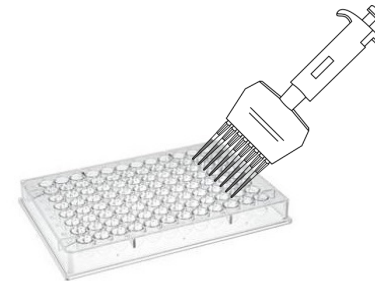


Add degrader molecule(s)



37°C
Incubate for 5-72 h

Add cell lysis reagent and detection reagent containing complementing EA



Incubate for 1 h

Read signal on plate reader



Building Biosensors for BRD4-Responsive Cancer Models

- Treatment with BET inhibitors (BETi) leads to decreases in c-Myc expression
- Several BETi (JQ1, I-BET-151, OTX015) with preclinical and clinical data supporting therapeutic use
- PROTACs targeting degradation of BRD4 also lead to decrease in c-Myc expression
- BETi and PROTACs to BRD4 are commercially available

Cell Models

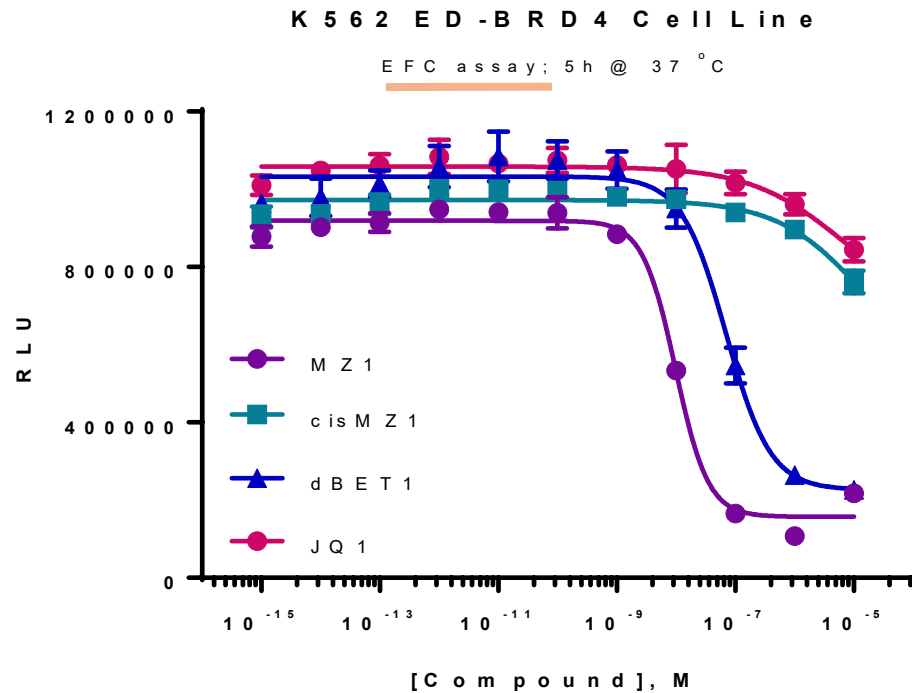
Cell lines	Origins	BRD4	c-Myc
K-562	Chronic myeloid leukemia (CML) cell line	++	+++
HCT116	Colorectal carcinoma cell line	+++	++++

Tool Molecules

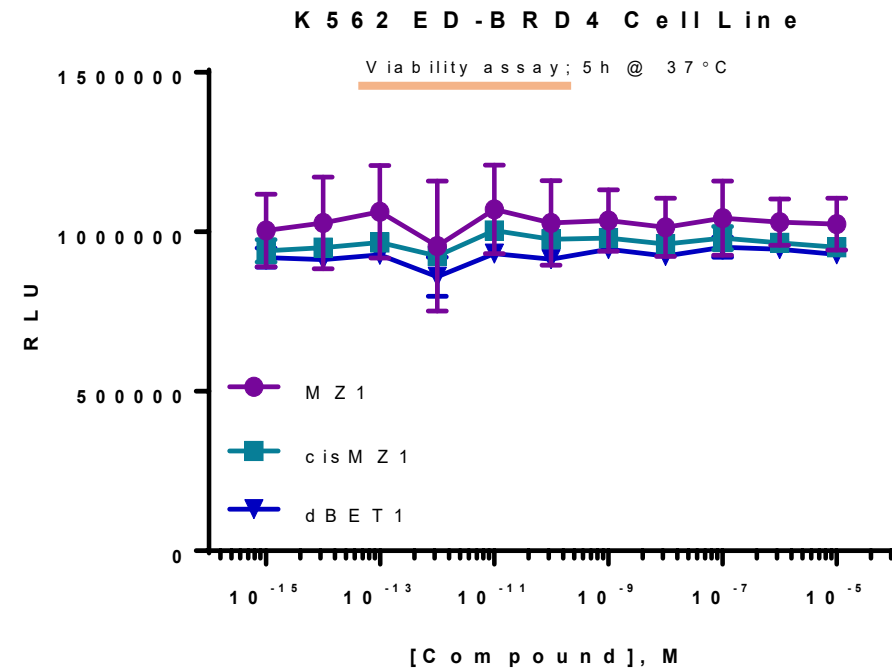
Tool Molecules	BRD4 Ligand	E3 Ligase Ligand
MZ1*	JQ1	VHL-1 (VHL)
dBET1*	JQ1	Thalidomide (cereblon)
cisMZ1*	JQ1	--
JQ1	JQ1	N/A

*PROTAC molecules

Rapid Degradation of ED-BRD4 in K562 Cells with BRD4 Targeting Agents



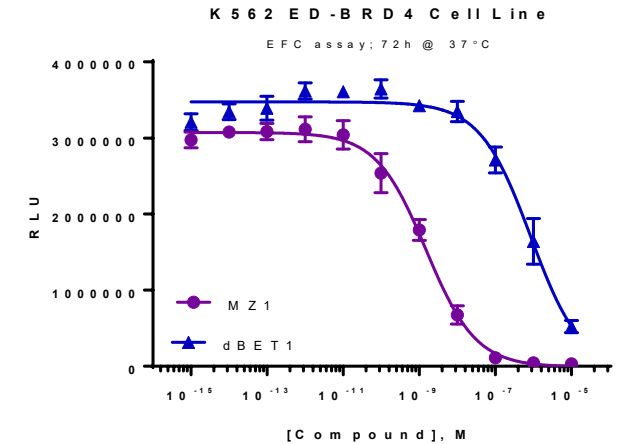
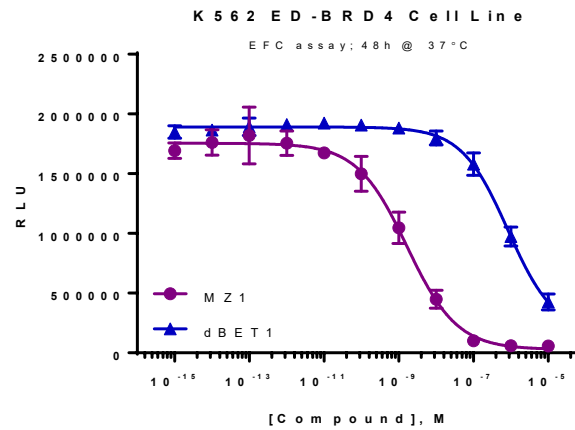
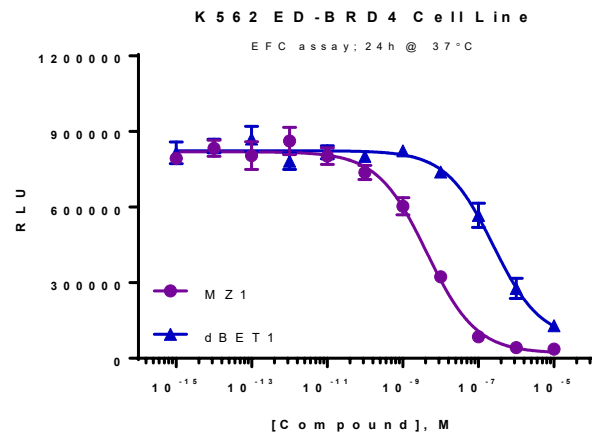
EFC Assay (functional)



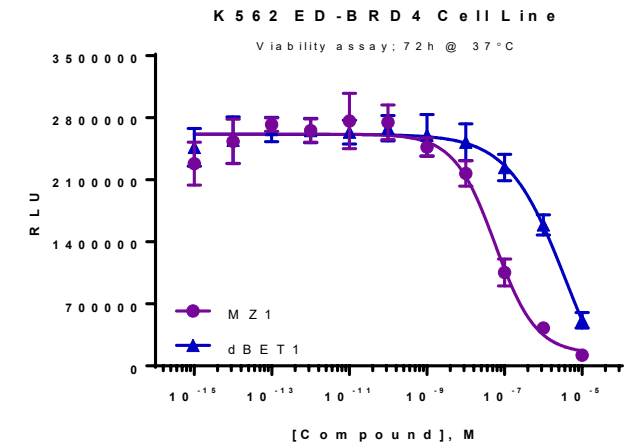
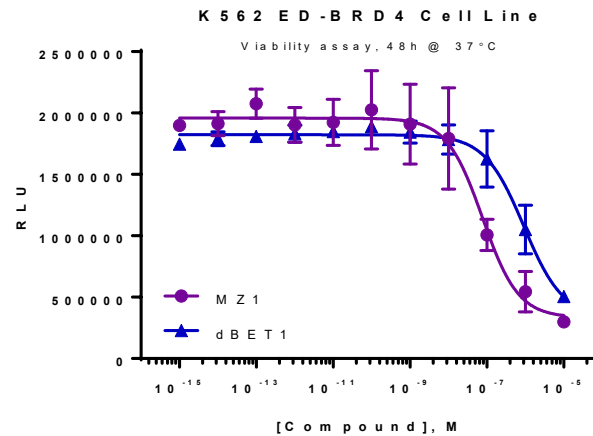
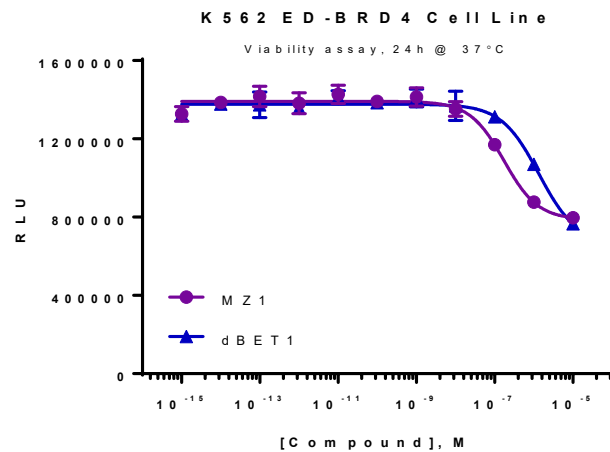
Phenotypic Assay (cell viability)

Time Course for MZ1 and dBET1 Mediated Degradation of ED-BRD4 in K562 Cells and Cell Viability

EFC Assays (24, 48 & 72 hours)



Viability Assays (24, 48 & 72 hours)



MZ1 Mediated Degradation of ED-BRD4 and Turnover of ED-c-Myc in K562 Cells

EFC Assay (functional)

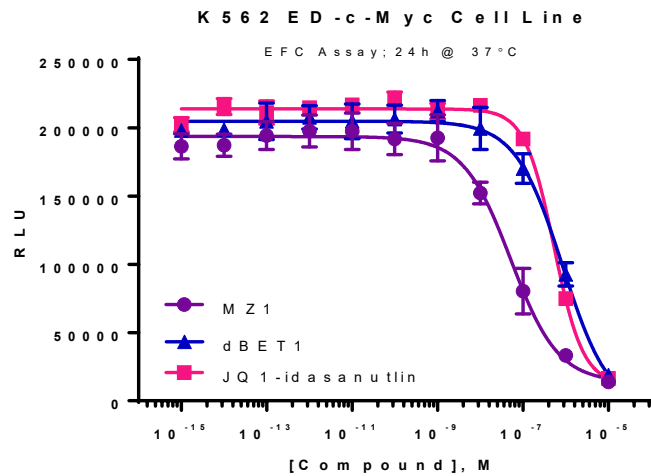
Biosensor	ED-BRD4 (K562)		ED-c-Myc (K562)		Fold Difference
	EC ₅₀ , nM	S/B	EC ₅₀ , nM	S/B	EC ₅₀ , nM
5h	9.7	5.8	222	15	22.9
24h	4.3	18	51	14.3	11.9
48h	1.6	28	27	32	16.9
72h	1.5	66	13	14	8.7

Phenotypic Assay (cell viability)

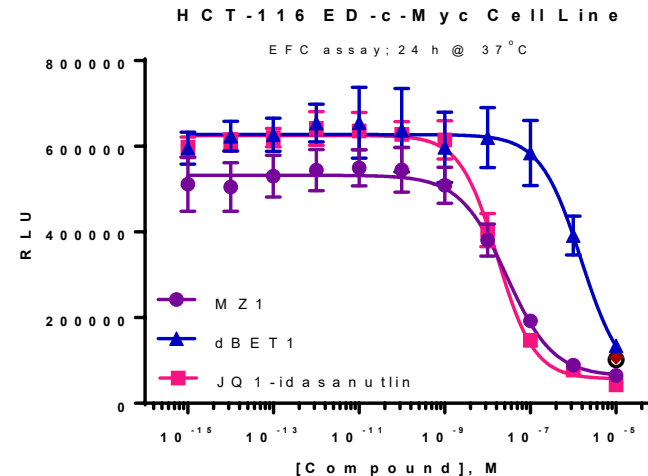
ED-BRD4 (K562)	
EC ₅₀ , nM	S/B
--	--
172	1.6
77	6.3
57	19

EFC-based biosensor cells are fast and sensitive

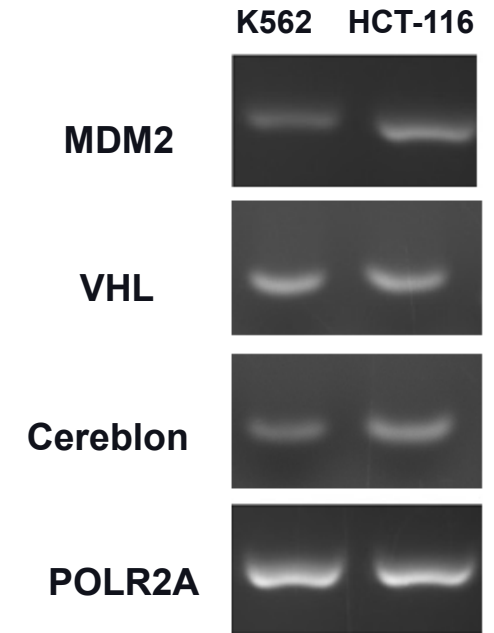
Differential PROTAC Efficacies Among Cell Models Correlates with Relative E3 Ligase Expression



MZ1 > dBET1 > JQ1-idasanutlin



MZ1 > JQ1-idasanutlin > dBET1



Relevant cell model makes a difference

PROTAC	BRD4 ligand	E3 ligase ligand
JQ1-idasanutlin	JQ1	idasanutlin (MDM2)
MZ1	JQ1	VHL-1 (VHL)
dBET1	JQ1	Thalidomide (cereblon)

Cellular EFC Biosensor Cell Lines to Quantify Protein Turnover

- We have generated an **EFC-based assay platform** to quantify changes in **endogenous** protein levels in disease relevant cell models
- As a proof-of-concept, we have applied this assay platform for detection of drug-induced changes in endogenous **BRD4** levels and its downstream target, **c-Myc** in blood cancer (K-562) and colon cancer (HCT-116) cell models
- The **high sensitivity** of our assay platform allows the detection of target protein turnover induced by targeted degrader molecules, such as PROTACs, with more rapid kinetics than phenotypic endpoint assays (e.g. cell proliferation)

These SPRINTer™ biosensor cell lines provide a **screening platform** to identify new molecular entities that modulate oncogenic protein levels for therapeutic development

SPRINTer Protein Turnover Biosensor Cell Lines
K562 BRD4 Cell Line
K562 c-Myc Cell Line
HCT-116 BRD4 Cell Line
HCT-116 c-Myc Cell Line

Visit discoverx.com/turnover to learn more

Thank You

Ksenya Cohen Katsenelson, Ph.D.
Group Leader, R&D, San Diego

Chao-Tsung Yang, Ph.D.
Principal Scientist, R&D, Fremont