

## PRODUCT DATASHEET

**Ready-to-Assay™ CX<sub>3</sub>CR1 Chemokine  
Receptor Frozen Cells****CATALOG NUMBER: HTS015RTA****Lot: 22F1304****CONTENTS:** Pack contains 2 vials of mycoplasma-free cells, 1 ml per vial. Fifty (50) mL of Media Component.**STORAGE:** Vials are to be stored in liquid N<sub>2</sub>. Media Component at 4°C (-20°C for prolonged storage).**BACKGROUND**

Ready-to-Assay GPCR frozen cells are designed for simple, rapid calcium assays with no requirement for intensive cell culturing. Eurofins Discovery Services has optimized the freezing conditions to provide cells with high viability and functionality post-thaw. The user simply thaws the cells and resuspends them in media, dispenses cell suspension into assay plates and, following over night recovery, assays for calcium response.

CX<sub>3</sub>CR1 is a GPCR expressed on natural killer cells, cytotoxic T lymphocytes, and macrophages. The sole ligand for CX<sub>3</sub>CR1, fractalkine, is an unusual chemokine that is expressed as a transmembrane molecule with a CX<sub>3</sub>C domain and a mucin domain (Imai *et al.*, 1997). Fractalkine is highly expressed on endothelial cells activated by TNF $\alpha$  and other proinflammatory cytokines, and fractalkine/CX<sub>3</sub>CR1 interactions mediate recruitment of macrophages into the atherosclerotic plaque (Lesnick *et al.*, 2003; McDermott *et al.*, 2003). In addition, fractalkine and CX<sub>3</sub>CR1 have been implicated in the pathogenesis of glomerulonephritis, HIV infection and rheumatoid arthritis (Ito *et al.*, 2002; Faure *et al.*, 2003; Nanki *et al.*, 2002). Cloned human CX<sub>3</sub>CR1-expressing cell line is made in the Chem-4 host, which supports high levels of recombinant CX<sub>3</sub>CR1 expression on the cell surface and contains optimized levels of a recombinant promiscuous G protein to couple the receptor to the calcium signaling pathway. Thus, the cell line is an ideal tool for screening for agonists, antagonists and modulators at CX<sub>3</sub>CR1.

**USE RESTRICTIONS**

Please see User Agreement (Label License) for further details. ***One such restriction is that the contents of the supplied vial(s) are limited to a single use and shall not be propagated and/or re-frozen by licensee.***

**WARNINGS**

For Research Use Only; Not for Use in Diagnostic Procedures  
Not for Animal or Human Consumption

**GMO**

This product contains genetically modified organisms.  
Este producto contiene organismos genéticamente modificados.  
Questo prodotto contiene degli organismi geneticamente modificati.  
Dieses Produkt enthält genetisch modifizierte Organismen.  
Ce produit contient organismes génétiquement des modifiés.  
Dit product bevat genetisch gewijzigde organismen.  
Tämä tuote sisältää geneettisesti muutettuja organismeja.  
Denna produkt innehåller genetiskt ändrade organismer.

## APPLICATIONS

Calcium Flux Assays

### APPLICATION DATA

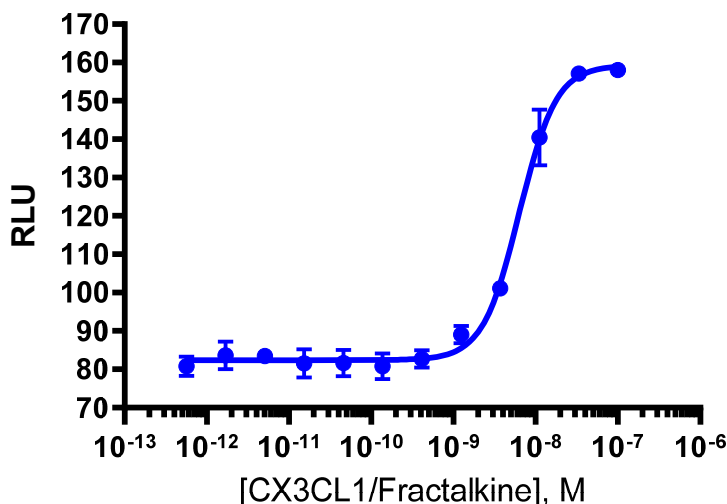


Figure 1. Representative data for activation of CX<sub>3</sub>CR1 receptor. Calcium flux in CX<sub>3</sub>CR1-expressing Chem-4 cell line induced by Fractalkine. CX<sub>3</sub>CR1-expressing Chem-4 cells were loaded with a calcium dye, and calcium flux in response to the indicated ligand(s), 3-fold serial dilution with each concentration performed in duplicate, was determined on a Molecular Devices FLIPR<sup>TETRA</sup>.

Table 1. Comparison of EC<sub>50</sub> values of CX<sub>3</sub>CR1-expressing Chem-4 cells with values described in the literature.

LIGAND	ASSAY	POTENCY (nM)	REFERENCE
Fractalkine	Calcium Flux	6.3	Eurofins Internal Data

## ASSAY SETUP

1. Immediately upon receipt, thaw cells or place cells in liquid nitrogen.
2. Thaw cells rapidly by removing from liquid nitrogen and immediately immersing in a 37°C water bath. Immediately after ice has thawed, sterilize the exterior of the vial with 70% ethanol.
3. Add 1mL of pre-warmed Media Component to each vial of cells. Place contents from two vials into a 15 mL conical tube and bring the volume to 10 mL of Media Component.
4. Centrifuge the cell suspension at 190 x g for four minutes
5. Remove supernatant and add 10.5 mL of pre-warmed Media Component to resuspend the cell pellet.
6. Seed cell suspension into appropriate assay microplate (100 µL/well for 96-well plate, 25 µL/well for 384-well plate).
7. When seeding is complete, place the assay plate at room temperature for 30 minutes.
8. Move assay plate to a humidified 37°C 5% CO<sub>2</sub> incubator for 24 hours.
9. After 24 hour incubation, remove assay plate from the incubator and invert plate to remove all Media Component.
10. Prepare Fluo-8, AM (AAT Bioquest: 21080) Ca<sup>2+</sup> dye by dissolving 1mg of Fluo-8 NW in 200 µL of DMSO.

Once dissolved place 10  $\mu\text{L}$  of Fluo-8 NW  $\text{Ca}^{2+}$  dye solution into 10 mL of HBSS 20mM HEPES, 2.5mM Probenecid pH 7.4 buffer and apply to assay microplate ( $\text{Ca}^{2+}$  dye at 10  $\mu\text{L}$  /10 mL is sufficient for loading one (1) microplate).

11. Set-up FLIPR to dispense 3x ligand to appropriate wells in the assay plate. Set excitation wavelength at 470-495 nm (FLIPR<sup>TETRA</sup>) or 485 nm (FLIPR1, FLIPR2, FLIPR3) and emission wavelength at 515-565 nm (FLIPR<sup>TETRA</sup>) or emission filter for  $\text{Ca}^{2+}$  dyes (FLIPR1, FLIPR2, FLIPR3). Set pipet tip height to 5  $\mu\text{L}$  below liquid level and dispense rate to 75  $\mu\text{L}/\text{sec}$  (96-well format) or 50  $\mu\text{L}/\text{sec}$  (384-well format). Set up plate layout and tip layout for each individual experiment. Set time course for 180 seconds, with ligand addition at 10 seconds.
12. Ligands are prepared in non-binding surface Corning plates (Corning 3605 – 96-well or Corning 3574 – 384-well).
13. After the run is complete, negative control correction is applied and data analyzed utilizing the maximum statistic.

## ASSAY MATERIALS

### Description

### Supplier and Product Number

HBSS	Hyclone: SH3026802
HEPES 1M Stock	EMD Millipore: TMS-003-C
Probenecid	Sigma: P8761
Quest Fluo-8 <sup>TM</sup> , AM	AAT Bioquest: 21080
Fractalkine	Peprotech: 300-31
Non-binding white plates (for ligand prep)	Corning: 3605(96-well)/3574(384-well)
Black (clear bottom) tissue-culture treated plates	Corning: 3904(96-well)/3712(384-well)

## FLIPR SETTINGS

Settings for FLIPR<sup>TETRA</sup>® with ICCD camera option

### Option

### Setting

Read Mode	Fluorescence
Ex/Em	Ex470_495 / Em515_575
Camera Gain	2000
Gate Open	6 %
Exposure Time	0.53
Read Interval	1s
Dispense Volume	50 $\mu\text{L}$ (25 $\mu\text{L}$ for 384-well)
Dispense Height	25 $\mu\text{L}$ (50 $\mu\text{L}$ for 384-well)
Dispense Speed	75 $\mu\text{L}$ L/sec (50 $\mu\text{L}$ for 384-well)
Expel Volume	0 $\mu\text{L}$
Analysis	Subtract Bias Sample 1

## HOST CELL

Chem-4, an adherent rat hematopoietic cell line expressing endogenous  $\text{G}\alpha_{15}$  protein as well as an exogenous proprietary promiscuous  $\text{G}\alpha$  protein.

## EXONGENOUS GENE EXPRESSION

CX<sub>3</sub>CR1cDNA (Accession Number: U28934; see CODING SEQUENCE below) expressed from a proprietary plasmid.

## RELATED PRODUCTS

PRODUCT NUMBER	DESCRIPTION
<b>HTSCHEM-1RTA</b>	Ready-to-Assay™ Chem-1 host frozen cells (control cells)
<b>HTS160M</b>	ChemiScreen™ CX <sub>3</sub> CR1 Chemokine receptor membrane prep

\* Note: Chem-4 cells are derived from Chem-1 cells

## REFERENCES

1. Faure S. *et al.* (2003) Deleterious genetic influence of CX<sub>3</sub>CR1 genotypes on HIV-1 disease progression. *J Acquir. Immune Defic. Syndr.* 32: 335-7
2. Imai T. *et al.* (1997) Identification and molecular characterization of fractalkine receptor CX<sub>3</sub>CR1, which mediates both leukocyte migration and adhesion. *Cell* 91: 521-30.
3. Ito Y. *et al.* (2002) Fractalkine expression and the recruitment of CX<sub>3</sub>CR1+ cells in the prolonged mesangial proliferative glomerulonephritis. *Kidney Int.* 61: 2044-57.
4. Lesnick P. *et al.* (2003) Decreased atherosclerosis in CX<sub>3</sub>CR1-/- mice reveals a role for fractalkine in atherogenesis. *J. Clin. Invest.* 111: 333-40.
5. McDermott D.H. *et al.* (2003) Chemokine receptor mutant CX<sub>3</sub>CR1-M280 has impaired adhesive function and correlates with protection from cardiovascular disease in humans. *J. Clin. Invest.* 111: 1241-50.
6. Nanki T. *et al.* (2002) Migration of CX<sub>3</sub>CR1-positive T cells producing type 1 cytokines and cytotoxic molecules into the synovium of patients with rheumatoid arthritis. *Arthritis Rheum.* 46: 2878-83.

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