

## PRODUCT DATASHEET

### Ready-to-Assay™ CXCR1 Chemokine Receptor Frozen Cells

#### CATALOG NUMBER: HTS001RTA

**CONTENTS:** Pack contains 2 vials of mycoplasma-free cells, 1 ml per vial. Fifty (50) mL of Media Component.

**STORAGE:** Vials are to be stored in liquid N<sub>2</sub>. Media Component at 4°C (-20°C for prolonged storage).

#### BACKGROUND

Ready-to-Assay™ GPCR frozen cells are designed for simple, rapid calcium assays with no requirement for intensive cell culturing. Eurofins Discovery Services has optimized the freezing conditions to provide cells with high viability and functionality post-thaw. The user simply thaws the cells and resuspends them in media, dispenses cell suspension into assay plates and, following overnight recovery, assays for calcium response.

CXCR1 (also known as IL-8RA) is a Gi/o-coupled GPCR expressed on neutrophils, CD8(+) T cells, and intestinal epithelial cells (Murphy, 1997; Sturm et al., 2005; Takata et al., 2004). CXCR1 binds with high affinity to the chemokine interleukin-8 (IL-8) and much lower affinity to GRO $\alpha$ , whereas a related receptor, CXCR2, binds with equal affinity to IL-8 and GRO $\alpha$  (Hall et al., 1999). CXCR1 has been proposed to be the primary regulator of neutrophil response to IL-8 in sepsis (Cummings et al., 1999). Antagonists of CXCR1 and CXCR2 have been developed and display efficacy in animal models of reperfusion injury (Bertini, et al., 2004; Kaneider et al., 2005). Cloned human CXCR1-expressing cell line is made in the Chem-1 host, which supports high levels of recombinant CXCR1 expression on the cell surface and contains high levels of the promiscuous G protein G $\alpha$ 15 to couple the receptor to the calcium signaling pathway. Thus, the cell line is an ideal tool for screening for agonists, antagonists and modulators at CXCR1.

#### USE RESTRICTIONS

Please see User Agreement (Label License) for further details. ***One such restriction is that the contents of the supplied vial(s) are limited to a single use and shall not be propagated and/or re-frozen by licensee.***

#### WARNINGS

For Research Use Only; Not for Use in Diagnostic Procedures  
Not for Animal or Human Consumption

#### GMO

This product contains genetically modified organisms.  
Este producto contiene organismos genéticamente modificados.  
Questo prodotto contiene degli organismi geneticamente modificati.  
Dieses Produkt enthält genetisch modifizierte Organismen.  
Ce produit contient organismes génétiquement des modifiés.  
Dit product bevat genetisch gewijzigde organismen.  
Tämä tuote sisältää geneettisesti muutettuja organismeja.  
Denna produkt innehåller genetiskt ändrade organismer.

## APPLICATIONS

Calcium Flux Assays

### APPLICATION DATA

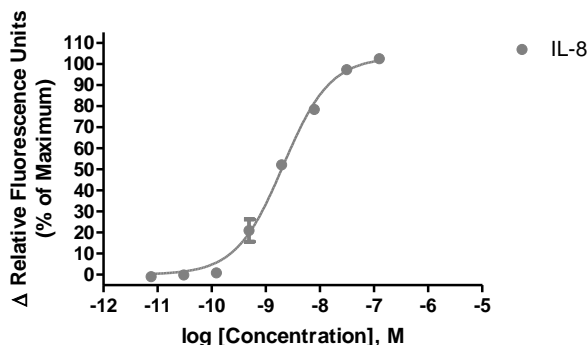


Figure 1. Representative data for activation of CXCR1 receptor. Calcium flux in CXCR1-expressing Chem-1 cell line induced by IL-8 (Interleukin 8). CXCR1-expressing Chem-1 cells were loaded with a calcium dye, and calcium flux in response to the indicated ligand(s), 4-fold serial dilution with each concentration performed in duplicate, was determined on a Molecular Devices FLIPR<sup>TETRA</sup> with ICCD camera. Maximal fluorescence signal obtained in this experiment was 17,800 RLU (Relative Light Units).

Table 1. Comparison of EC<sub>50</sub> values of CXCR1-expressing Chem-1 cells with values described in the literature.

LIGAND	ASSAY	POTENCY (nM)	REFERENCE
IL-8	Calcium Flux	2.1	Eurofins Internal Data

## ASSAY SETUP

1. Immediately upon receipt, thaw cells or place cells in liquid nitrogen.
2. Thaw cells rapidly by removing from liquid nitrogen and immediately immersing in a 37°C water bath. Immediately after ice has thawed, sterilize the exterior of the vial with 70% ethanol.
3. Add 1mL of pre-warmed Media Component to each vial of cells. Place contents from two vials into a 15 mL conical tube and bring the volume to 10 mL of Media Component.
4. Centrifuge the cell suspension at 190 x g for four minutes
5. Remove supernatant and add 10.5 mL of pre-warmed Media Component to resuspend the cell pellet.
6. Seed cell suspension into appropriate assay microplate (100 µL/well for 96-well plate, 25 µL/well for 384-well plate).
7. When seeding is complete, place the assay plate at room temperature for 30 minutes.
8. Move assay plate to a humidified 37°C 5% CO<sub>2</sub> incubator for 24 hours.
9. After 24 hour incubation, remove assay plate from the incubator and wash sufficiently with Hank's Balanced Salt Solution (HBSS) supplemented with 20mM HEPES, 2.5mM Probenecid at pH 7.4 to remove all trace of Media Component.

10. Prepare Fluo-8, AM (AAT Bioquest: 21080) Ca<sup>2+</sup> dye by dissolving 1mg of Fluo-8 NW in 200 µL of DMSO. Once dissolved place 10 µL of Fluo-8 NW Ca<sup>2+</sup> dye solution into 10 mL of HBSS 20mM HEPES, 2.5mM Probenecid pH 7.4 buffer and apply to assay microplate (Ca<sup>2+</sup> dye at 10 µL /10 mL is sufficient for loading one (1) microplate).
11. Set-up FLIPR to dispense 3x ligand to appropriate wells in the assay plate. Set excitation wavelength at 470-495 nm (FLIPR<sup>TETRA</sup>) or 485 nm (FLIPR1, FLIPR2, FLIPR3) and emission wavelength at 515-565 nm (FLIPR<sup>TETRA</sup>) or emission filter for Ca<sup>2+</sup> dyes (FLIPR1, FLIPR2, FLIPR3). Set pipet tip height to 5 µL below liquid level and dispense rate to 75 µL/sec (96-well format) or 50 µL/sec (384-well format). Set up plate layout and tip layout for each individual experiment. Set time course for 180 seconds, with ligand addition at 10 seconds.
12. Ligands are prepared in non-binding surface Corning plates (Corning 3605 – 96-well or Corning 3574 – 384-well).
13. After the run is complete, negative control correction is applied and data analyzed utilizing the maximum statistic.

## ASSAY MATERIALS

Description	Supplier and Product Number
HBSS	Hyclone: SH30268.02
HEPES 1M Stock	EMD Millipore.: TMS-003-C
Probenecid	Sigma: P8761
Quest Fluo-8™, AM	AAT Bioquest: 21080
IL-8 ligand	Chemicon: IL008
Non-binding white plates (for ligand prep)	Corning: 3605(96-well)/3574(384-well)
Black (clear bottom) tissue-culture treated plates	Corning: 3904(96-well)/3712(384-well)

## FLIPR SETTINGS

Settings for FLIPR<sup>TETRA</sup>® with ICCD camera option

Option	Setting
Read Mode	Fluorescence
Ex/Em	Ex470_495 / Em515_575
Camera Gain	2000
Gate Open	6 %
Exposure Time	0.53
Read Interval	1s
Dispense Volume	50 µl (25 µl for 384-well)
Dispense Height	25 µl (50 µl for 384-well)
Dispense Speed	75 µl L/sec (50 µl for 384-well)
Expel Volume	0 µl
Analysis	Subtract Bias Sample 1

## HOST CELL

Chem-1, an adherent rat hematopoietic cell line expressing endogenous Gα15 protein.

## EXONGENOUS GENE EXPRESSION

IL8RA cDNA (Accession Number: M68932) expressed from a proprietary pHS plasmid.

## RELATED PRODUCTS

PRODUCT NUMBER	DESCRIPTION
<b>HTSCHEM-1RTA</b>	Ready-to-Assay™ Chem-1 host frozen cells (control cells)
<b>HTS001C</b>	ChemiScreen™ CXCR1 Chemokine receptor stable cell line
<b>HTS001M</b>	ChemiScreen™ CXCR1 Chemokine receptor membrane prep

## REFERENCES

1. Bertini R et al. (2004) Noncompetitive allosteric inhibitors of the inflammatory chemokine receptors CXCR1 and CXCR2: prevention of reperfusion injury. *Proc. Natl. Acad. Sci. U S A.* 101(32):11791-6.
2. Cummings CJ et al. (1999) Expression and function of the chemokine receptors CXCR1 and CXCR2 in sepsis. *J. Immunol.* 162(4):2341-6.
3. Hall DA et al. (1999) Signalling by CXC-chemokine receptors 1 and 2 expressed in CHO cells: A comparison of calcium mobilization, inhibition of adenylyl cyclase and stimulation of GTP•S binding induced by IL-8 and GROa. *Br. J. Pharmacol.* 126: 810-818.
4. Kaneider NC et al. (2005) Reversing systemic inflammatory response syndrome with chemokine receptor pepducins. *Nat. Med.* 11(6):661-5.
5. Murphy PM (1997) Neutrophil receptors for interleukin-8 and related CXC chemokines. *Semin. Hematol.* 34: 311-8.
6. Sturm A et al. (2005) CXCL8 modulates human intestinal epithelial cells through a CXCR1 dependent pathway. *Cytokine* 29(1):42-8.
7. Takata H et al. (2004) Cutting edge: expression of chemokine receptor CXCR1 on human effector CD8+ T cells. *J. Immunol.* 173(4):2231-5.

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