Simple, Reproducible & Robust Bioassays In Cryopreserved Ready-To-Assay Format

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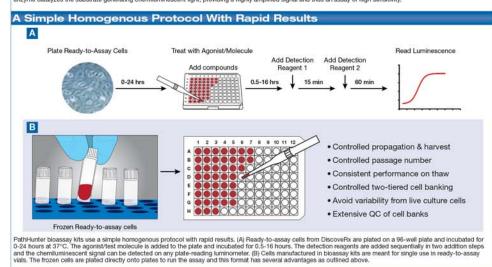
Abstract

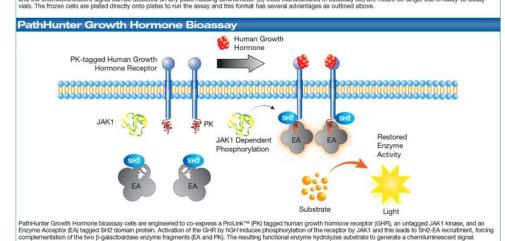
Cell-based bioassays often pose a hurdle during a rapidly moving biologics development program. High standards for assay accuracy, precision, reproducibility and robustness are additionally put to the test by the use of continuous culture cells that can add to assay variability and increase the cost and complexity of each assay.

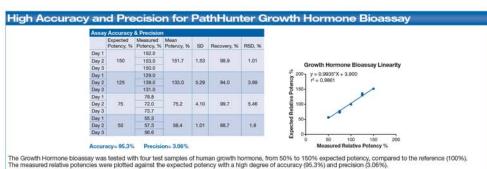
Advent of new technology platforms can enable rapid development of cell-based bioassays that are simple, accurate, precise and robust. Here, we discuss the development and application of diverse PathHunter® cell-based assays that cover distinct cellular mechanisms either distal from or proximal to the receptor. These quantitative assays are highly specific, scalable, robust and utilize a homogenous mix-and-read protocol, which facilitates rapid and reproducible detection of drug potency. In order to increase the reproducibility and reduce the complexity of the assay by eliminating cell culture, these assays are developed as cryopreserved ready-to-assay cells that can be plated directly from the frozen state onto the assay plate. The cell preparation, bioassay protocol and reagents have been optimized to provide superior bi performance with high reproducibility (<10%RSD). Examples discussed here include assays for Bevacizumab, human growth hormone, GLP

PathHunter Enzyme Fragment Complementation

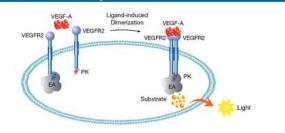
DiscoveRx's proprietary PathHunter Enzyme Fragment Complementation (EFC) technology consists of the β-galactosidase (β-gal) enzyme, split into two inactive complements, the enzyme donor peptide (ED) and enzyme acceptor (EA). When brought together in close proximity, ED complements with EA forming active β-gal. The active enzyme catalyzes the substrate generating chemiliuminescent light, providing a highly ampfilled signal must have an easily of high sensitivity.







PathHunter Bevacizumab Bioassay



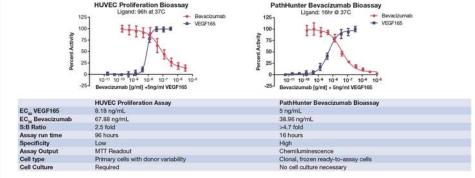
Using EFC, we have generated a simple cell-based assay for VEGF-A and the VEGFR2 receptor. The assay measures VEGFR2 receptor homodimerization by VEGF-A as the first step in its activation cascade that eventually leads to angiogenesis. The VEGFR2 receptor is tagged with both the ProLink (PK) and the Enzyme Acceptor (EA). Upon ligand induced activation, receptor dimerization occurs which forces the two b-gal components to complement. This creates an active enzyme that hydrolyzes substrate exceptions as hereally improved the state.

cellent Accuracy and Precision for PathHunter Bevacizumab Bioassay

	Expected Potency, %	Measured Potency, %	Mean Potency, %	SD	Recovery, %	RSD, %
lay 1		151.0				
ay 2	150	144.0	148.7	4.04	99,1	5.12
у 3		151.0				
ay 1		109.0				
xy 2	125	115.0	113.0 3	3.46	6 90.4	3.07
ay 3		115.0				
ay.1		74.4				
sy 2	75	73.9	72.7	2.47	97.0	3.12
y 3		69.9				
ay 1		48.8				
lay 2	50	45.9	48.5	2.42	96.9	4.99
lay 3		50.7				

VEGF-A is known to cause homodimerization of VEGFR2 (KDR), as the first step in the activation cascade of these receptors. Anti-VEGF-A antibodies such as Avastin (Bevac zumab), Ranibizumab and Aflibercept bind to VEGF-A and prevent this dimer formation, leading to inhibition of VEGF-A dependent signaling. Here we have tested the VEG-FR2 homodimer assay with VEGF-A, demonstrating a robust response and a high level of reproducibility with multiple runs.

The VEGFR dimerization assay was tested with four test samples, from 50% to 150%, compared to a reference standard (100%). The measured relative potencies were plotted against the expected relative potencies with a very high degree of accuracy and precision.

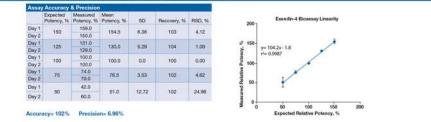


The HUVEC proliferation assay is another method to test drug potency for the anti-VEGF drugs. VEGF-A (VEGF165 is an active splice variant of VEGF-A) will cause HUVEC cells to proliferate and Bevacizumab will inhibit this by inhibiting VEGF-A interaction with VEGF-R2. Here we have tested the Path-Hunter Bevacizumab biosassay with VEGF-A and bevacizumab floor gight, demonstrating the robust and precise response to both agents, and EC/CS comparable to those obtained in the HUVEC proliferation assay (top left). The table below compares the proliferation assay and the Path-Hunter assay, demonstrating that the latter is quicker, more robust, highly specific and generates the

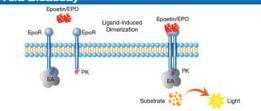
AMP Hunter Bioassay for GLP1R Agonists

as in intracellular cAMP levels in response to agonist stimulation of the GLP1 receptor, through GLP1/G-37) or Exendin-4. The cellular cAMP levels are detected using the cAMP detection kit, where cAMP from the cells and ED-labeled cAMP (ED-cAMP) compete for anti-cAMP artibody (Ab) binding sites. Antibody-bound ED-cAMP will not be able to complement with EA, but unbound ED-cAMP is free to complement EA to form active enzyme, which subsequently produces a luminescent signal. The amount of signal produced is directly proportional to the amount of cAMP in the cells.

high accuracy with Exendin-4 in the GLP1R Bioassay

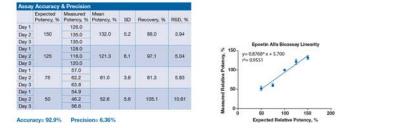


The bioassay for GLP1 recentor agonists was tested with four test samples of Exendin-4, from 50% to 150% expected potency, compared to the reference (100%). Each assay was run in triplicate on each day. The measured relative potencies were plotted against the expected potency with a high degree of accuracy (102%) and precision (6,96%).



The PathHunter Epoetin Alfa Bioassay is used to detect ligand induced dimerization of two subunits of a receptor-dimer pair. This assay relies on the native biology of the erythropoletin receptor (EpoR), where erythropoletin induces receptor homodimerization as the first step in the pathway activation cascade. The cells have been engineered to cooxpress EpoR itsued to PK, and another EpoR construct fused to PK, and another EpoR construct fused to EA. Binding of Epoetin alfa or darbepoetin afta to EpoR induces it to homodimerize, forcing complementation of the two enzyme fragments, PK and EA. This results in the formation of a functional enzyme that hydrolyzes a substrate to generate a chemiliumines-

Potency, Linearity, Precision & Accuracy of the Epoetin Alfa Bioassay



The Epoetin alfa bloassay was tested with four test samples of erythropoeitin, from 50% to 150% expected potency, compared to the reference (100%), Each assay was run in triplicate on each of the days. The measured relative potencies were plotted against the expected potency with a high degree of accuracy (92.9%) and precision (6.36%).

- . Cell-based assays with functional response based on drug mechanism of action & native biology
- Excellent potency and linearity with high accuracy and precision of the bioassay
- · Simple protocol with rapid results on any benchtop luminom
- · Highly reproducible and robust
- · Used for both NAb and potency bioassays
- 750+ cell-based assays & 30+ bioassays for biosimilars available
- Developed as cryopreserved ready-to-assay cells

Go to www.discoverx.com/biologics to see additional validation data and a full list of targets.

